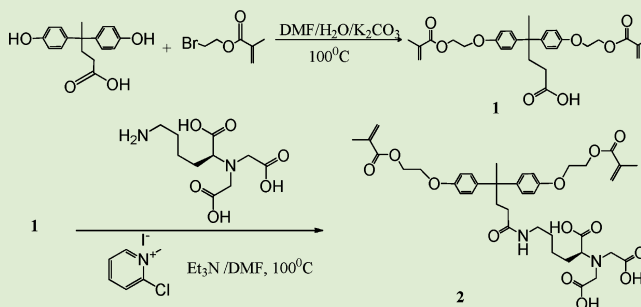


# Synthesis and Characterization of New Antibacterial Fluoride-Releasing Monomer and Dental Composite

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**ABSTRACT:** A new dimethacrylate chelating monomer containing a BisGMA-like backbone structure and a bis-(carboxymethyl)-L-lysine chelating group and its ternary zirconium–fluoride complex (antibacterial fluoride-releasing monomer) have been synthesized. The monomer structures were confirmed by  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, and ES-MS analysis. Several experimental fluoride-releasing dental composites containing different quantities of the new antibacterial fluoride-releasing monomer were formulated and tested for fluoride release, fluoride recharge, compressive and flexural strengths, water sorption, and solubility. These composites displayed high fluoride release and recharge capabilities, as well as good physical and mechanical properties.



Dental caries is still a major health problem worldwide. Resin-based dental composites (RBC) have been widely used to restore carious teeth, but they have limited service life (5–7 years). The leading cause for failure of RBCs is secondary (recurrent) caries.<sup>1–5</sup> Fluoride is well documented as the most effective and widely used anticariogenic agent. Fluoride-releasing dental materials may reduce secondary caries at the restoration's margins.<sup>6–9</sup> Current dental composites have high strength, good wear resistance, and excellent esthetics,<sup>10–12</sup> but their fluoride release rates and fluoride recharge capabilities are quite low.<sup>13</sup> The major challenge for the development of fluoride-releasing dental composites is to increase their fluoride-releasing and recharging capabilities while maintaining their physical and mechanical properties.

In recent years, our group has developed several novel fluoride-releasing dental monomers containing ternary zirconium–fluoride chelates and related dental composites.<sup>14–18</sup> In the early studies, dimethacrylate chelating monomers containing amino- or amido-diacetic acids and their ternary zirconium–fluoride chelates were synthesized. The experimental composites containing these novel fluoride-releasing monomers displayed significantly higher fluoride-release and recharge capabilities than commercial fluoride-releasing composites, but their physical and mechanical properties were significantly lower than commercial composites due to high water sorption and solubility.<sup>15,16</sup> Recently, the formulation of the fluoride-releasing composite was optimized to improve the physical and mechanical properties but its fluoride release was significantly reduced, though they were significantly higher than the conventional fluoride releasing dental composites.<sup>17</sup> Therefore, the major challenge for the development of fluoride-releasing dental composites remains to increase their fluoride-releasing and recharging capabilities while maintaining their physical and mechanical properties.

Here we report the synthesis of a new dimethacrylate chelating monomer containing bis(carboxymethyl)-L-lysine and its ternary zirconium–fluoride complex, the synthesis of the antibacterial fluoride-releasing monomer (AF), and the characterization of the experimental antibacterial fluoride-releasing dental composite containing such a monomer.

The new dimethacrylate chelating monomer **2** has been designed to have a BisGMA-like backbone structure and a tetra-dentate chelating group (three carboxylic acids and one amino group) (Scheme 1). Therefore, the resulting ternary zirconium–fluoride complex has higher stability than the monomers containing amino- or amido-diacetic acids previously reported.<sup>15,16</sup> The potential increase of hydrophilicity (water sorption) of this chelating group has been compensated by removal of the two pendent hydroxyl groups from the backbone structure. The calculated water–octanol partition coefficient  $C \log P = 4.43$  indicates the monomer is rather hydrophobic, which helps reduce water sorption of the composite.

The dimethacrylate chelating monomer **2** can be readily synthesized by two steps (Scheme 1). The monosubstitution was a side reaction in the first step. This directly affected the yield of intermediate **1**. By increasing the reaction temperature (100 °C) and increasing the amount of 2-bromoethyl methacrylate, the yield of intermediate **1** was increased to about 20%.

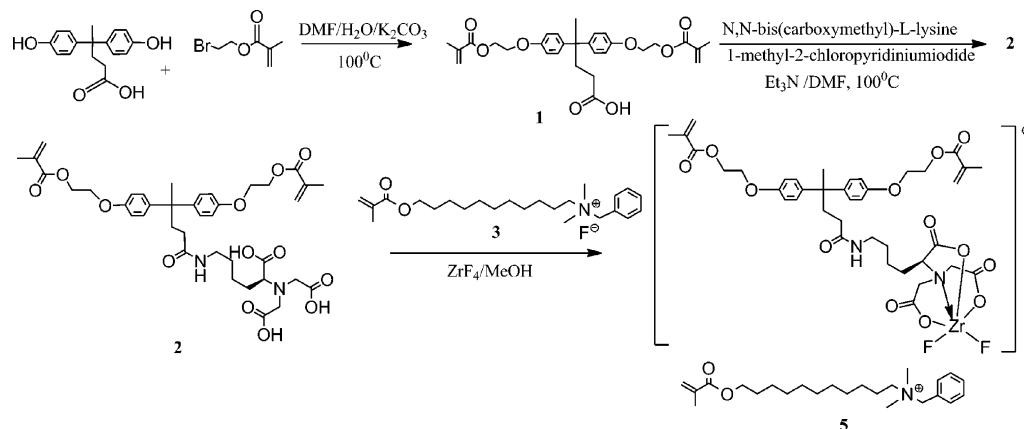
Chelating monomer **2** can react with  $\text{ZF}_4$ /methanol solution and **3** to form a ternary zirconium–fluoride complex **4** and antibacterial fluoride-releasing (AF) monomer **5**. The negative ion ES-MS spectra of monomers **2** and **5** are shown in Figures

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## Scheme 1. Synthesis of Chelating Monomer 2



1 and 2, respectively. The negative ion ES-MS spectrum of monomer 5 (Figure 2) shows high abundance of  $m/z = 879.0$

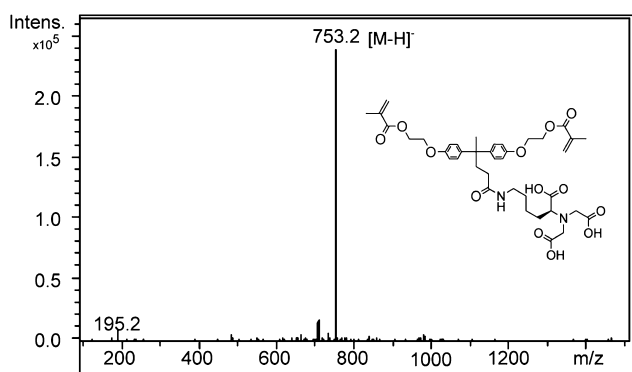


Figure 1. Negative ion ES-MS spectrum of chelating monomer 2.

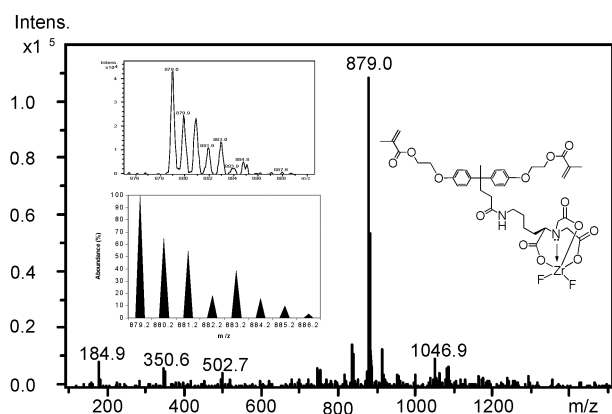


Figure 2. Negative ES-MS spectrum of the anion part of antibacterial fluoride releasing monomer 5. The insets are measured and calculated isotope distribution patterns of the peak  $m/z = 879.0$ .

and the unique isotope distribution pattern, which matches that of the anion part of the monomer  $[LZrF_2]^-$ . The AF monomer 5 includes a long-chain monomer [*N*-benzyl-11-(methacryloyloxy)]-*N,N*-dimethylundecan-1-ammonium] as the counterion. The positive ion ES-MS spectrum (not shown) of monomer 5 shows a single peak at  $m/z = 374.3051$  (calculated: 374.3054), which is the cation part of monomer 3. It has an antimicrobial effect, as reported in our previous study.<sup>19</sup>

Figure 3 shows the results of cytotoxicity of synthesized monomer and BisGMA at different concentrations. At  $10^{-5}$  M,

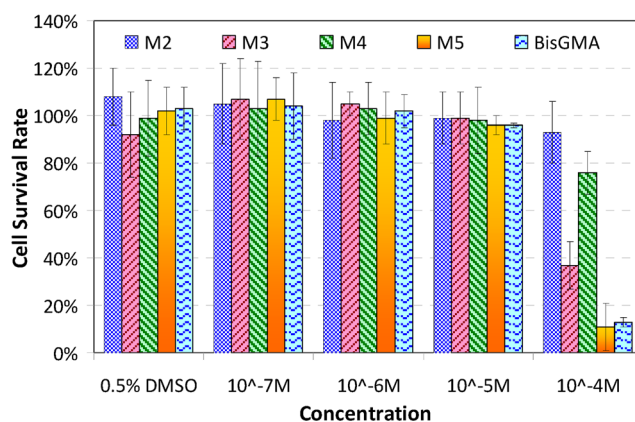


Figure 3. Cytotoxicity test: cell survival rate under different concentrations of monomers (M2, M3, M4, and M5 represents monomers 2, 3, 4, and 5, respectively).

all monomers show no cytotoxicity. At  $10^{-4}$  M, chelating monomer 2 had 93% cell survival rate, which indicates good biocompatibility. Monomer 4 had 73% cell survival rate, indicating slight cytotoxicity, which may have been caused by fluoride. Monomer 5 displayed some cytotoxicity (11% cell survival), but it is very similar to that of BisGMA (13%). Therefore, the synthesized monomers are acceptable for dental applications.

The experimental and control dental composites were formulated and their compositions are listed in Table 1. The flexural strength of the composites was tested after storage in

Table 1. Compositions of Experimental and Control Composites (wt %)

components	control	exp20	exp25	exp30
Bis GMA	11.6	8.7	6.525	6.09
EBPADMA	11.6	8.7	6.525	6.09
HDDMA	5.8	5.8	4.35	4.06
AF monomer	0	5.8	7.25	8.7
F-releasing filler	70	70	70	70
CQ	0.14	0.14	0.14	0.14
4E	0.58	0.58	0.58	0.58
PO	0.29	0.29	0.29	0.29

deionized water for 24 h and 3 months. Water sorption and solubility of the composites were also tested according ISO 4049–2000. The results are listed in Table 2. The experimental

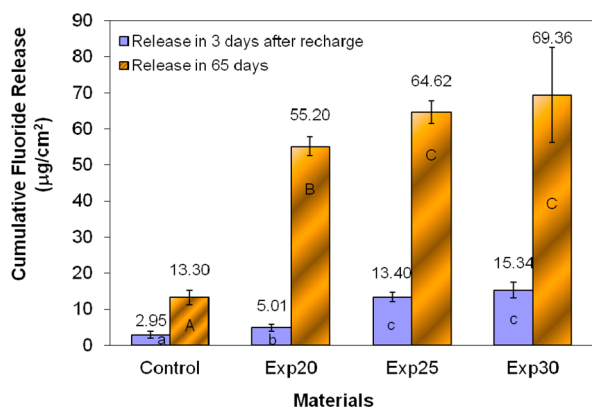
**Table 2. Physical and Mechanical Properties of Experimental and Control Composites<sup>a</sup>**

materials	flexure strength, 24 h (MPa)	flexure strength, 3 months (MPa)	water sorption ( $\mu\text{g}/\text{mm}^3$ )	solubility ( $\mu\text{g}/\text{mm}^3$ )
control	108.9 $\pm$ 9.7 <sup>A</sup>	96.1 $\pm$ 19.2 <sup>A</sup>	13.6 $\pm$ 1.2 <sup>a</sup>	1.48 $\pm$ 0.9 <sup>a</sup>
exp20	113.6 $\pm$ 15.9 <sup>A</sup>	80.8 $\pm$ 8.6 <sup>A,B</sup>	13.75 $\pm$ 1.1 <sup>a</sup>	1.84 $\pm$ 1.5 <sup>a</sup>
exp25	95.0 $\pm$ 11.9 <sup>A,B</sup>	80.6 $\pm$ 12.0 <sup>A,B</sup>	14.5 $\pm$ 2.1 <sup>ab</sup>	2.0 $\pm$ 0.8 <sup>a</sup>
exp30	85.1 $\pm$ 9.5 <sup>B</sup>	73.8 $\pm$ 7.9 <sup>B</sup>	20.5 $\pm$ 1.9 <sup>c</sup>	3.3 $\pm$ 0.4 <sup>b</sup>

<sup>a</sup>The groups with the same superscript letters in the same column have no significant difference ( $p > 0.05$ ).

composites containing up to 25% antibacterial fluoride-releasing (AF) monomer **5** in total monomers have similar physical and mechanical properties to the control composite ( $p = 0.244$ ). When the amount of AF monomer **5** is increased to 30% of the total monomers, the water sorption increases significantly and the mechanical properties decrease significantly.

Figure 4 shows the fluoride release and recharge profiles, cumulative fluoride release and recharge of the experimental



**Figure 4.** Cumulative fluoride release and fluoride recharge. The columns with the same letters have no significant difference ( $p > 0.05$ ).

and control composites. The experimental composite containing the new antibacterial fluoride-releasing monomer has significantly higher cumulative fluoride release and recharge capability than the control composite ( $p < 0.05$ ). In general, the fluoride release and recharge capabilities of the experimental composites increase with the increasing amount of AF monomer. However, a further increase in the amount of AF monomer above 25% has little effect on fluoride release and recharge, but it will increase water sorption and reduce mechanical properties of the composite. Therefore, the optimal concentration of AF monomer appears to be around 25% of the total monomers. In clinic application, such a new fluoride-releasing dental composite is expected to have a sustained fluoride release capability and enhanced caries-inhibitive effect. Therefore, it may prolong the service life of the composite restoration and reduce the cost for replacement.

In conclusion, the new dimethacrylate chelating monomer containing bis(carboxymethyl)-L-lysine and its ternary zirconium–fluoride complex (antibacterial fluoride-releasing mono-

mer) has been successfully synthesized. The experimental dental composite containing the new fluoride-releasing monomer displays high fluoride release and recharge capabilities as well as good physical and mechanical properties. The new monomer may be useful in the formulation of a variety of fluoride-releasing dental materials.

## EXPERIMENTAL METHODS

2-Bromoethyl methacrylate was purchased from Pfaltz and Bauer.  $N_{\omega}N_{\alpha}$ -bis(carboxymethyl)-L-lysine hydrate was purchased from Fluka. Bisphenol A dimethacrylate (BisGMA) was purchased from Polysciences. Ethoxylated bisphenol A dimethacrylate (EBPADMA) and 1,6-hexanediol dimethacrylate (HDDMA) were provided by Esstech. 4,4-Bis(4-hydroxyphenyl)valeric acid, 2-chloro-1-methyl-pyridinium iodide, camphorquinone (CQ), ethyl 4-dimethylaminobenzoate (4E), phenyl bis(2,4,6-trimethyl benzoyl)phosphine oxide (PO), and solvents were purchased from Aldrich. Silanized F-releasing glass filler was provided by Caulk/Dentsply. The antibacterial monomer **3** was synthesized using a previously reported method.<sup>19</sup> High resolution mass spectrometry using electrospray ionization (HRMS (ESI)) was carried out on Synapt HDMS (Waters). NMR spectra were acquired on Inova-400 (<sup>1</sup>H, <sup>13</sup>C) NMR spectrometer: <sup>1</sup>H at 400 MHz, <sup>13</sup>C at 125 MHz in CD<sub>3</sub>OD.

Synthesis of 4,4-bis[4-(2-methacryloyloxy)ethoxyphenyl]pentanoic acid (**1**): A 100 mL round-bottomed flask, equipped with a magnetic stirring bar was charged with 4,4-bis(4-hydroxyphenyl) valeric acid (5.72 g, 20 mmol), K<sub>2</sub>CO<sub>3</sub> (40 mmol), and DMF (60 mL). The mixture is stirred for 30 min. Bromoethyl methacrylate (9.57 g, 50 mmol) was added through one septum via a syringe over 10 min. The reaction mixture was stirred at bath temperature 40 °C for 48 h, and then additional bromoethyl methacrylate (10 mmol) was added at 100 °C and stirred for 24 h. After the reaction was completed, the reaction mixture was quenched by adding water (100 mL), and the pH was adjusted to 3 using HCl. The mixture was extracted with diethyl ether (3  $\times$  50 mL). The combined organic extracts were washed sequentially with brine (2  $\times$  50 mL), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure by a rotary evaporator. The crude product was purified on a silica gel column with CH<sub>3</sub>COOC<sub>2</sub>H<sub>5</sub>/hexane (3:2) as mobile phase to give (5.11 g, 50%) of compound **1**.

Analysis: <sup>1</sup>H NMR (CD<sub>3</sub>OD, 400 MHz)  $\delta$  1.54 (s, 3H), 1.93 (m, 6H), 2.13 (m, 2H), 2.38 (q, 2H), 4.17 (t, 2H), 4.30 (m, 4H), 2.47 (t, 2H), 5.58 (m, 2H), 6.12 (m, 2H), 6.73 (t, 2H), 6.80 (t, 2H), 7.02 (q, 2H), 7.08 (q, 2H); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 125 MHz)  $\delta$  18.47, 27.96, 30.42, 36.75, 44.70, 63.48, 66.11, 66.63, 114.39, 115.15, 126.47, 128.53, 136.11, 140.66, 156.74, 167.21, 174.25. HRMS (ESI) analysis (in MeOH/CH<sub>2</sub>Cl<sub>2</sub>, negative ion mode) for C<sub>29</sub>H<sub>45</sub>O<sub>8</sub>, [M – H]<sup>–</sup>,  $m/z$ : Found, 509.2207; Calcd, 509.2181.

Synthesis of (S)-2,2'-(5-(4,4-bis(4-(2-(methacryloyloxy)ethoxy)phenyl)pentanamido)-1-carboxypentylazanediyl)diacetic acid (**2**): A 250 mL round-bottomed flask equipped with a magnetic stirring bar was charged with 2-chloro-1-methyl-pyridinium iodide (2.81 g, 11 mmol) and DMF (100 mL). After it was dissolved, compound **1** (5.09 g, 10 mmol) was added. The reaction mixture was stirred at 100 °C for 2 h.  $N_{\omega}N_{\alpha}$ -bis(carboxymethyl)-L-lysine hydrate (2.88g, 11 mmol) was added. The reaction mixture was stirred vigorously for 18 h and quenched by adding water (100 mL). The pH value of the aqueous layer was adjusted to 3, and the solution was extracted with diethyl ether (3  $\times$  50 mL). The combined organic extracts were washed with brine (2  $\times$  150 mL), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure by a rotary evaporator. The crude product was purified on a silica gel column with CH<sub>3</sub>COOC<sub>2</sub>H<sub>5</sub>/CH<sub>3</sub>OH (1:4) as mobile phase to give (3.56 g, 47.2%) of compound **2**.

Analysis: <sup>1</sup>H NMR (CD<sub>3</sub>OD, 400 MHz)  $\delta$  1.29 (m, 2H), 1.43–1.62 (m, 4H), 1.89 (s, 3H), 1.96 (s, 6H), 2.13–2.35 (m, 4H), 2.98–3.10 (m, 6H), 3.85 (m, 1H), 4.19–4.56 (m, 8H), 5.58 (m, 2H), 6.16 (m, 2H), 6.73 (t, 2H), 6.80 (t, 2H), 7.02 (q, 2H), 7.08 (q, 2H), 8.01 (s, NH). <sup>13</sup>C NMR (CD<sub>3</sub>OD, 125 MHz)  $\delta$  18.48, 27.97, 30.40, 31.81,

36.80, 44.66, 55.31, 62.42, 63.64, 66.64, 114.35, 126.47, 128.53, 136.15, 139.33, 140.60, 140.85, 154.34, 167.67, 174.11, 174.22. HRMS (ESI; negative ion mode) for  $C_{39}H_{49}N_2O_{13}$   $[M - H]^-$ ,  $m/z$ : Found, 753.3303; Calcd, 753.3240.

Synthesis of fluoride-releasing monomer (4) and antibacterial fluoride-releasing monomer (5): Compound 2 (2 mmol) and  $ZrF_4/MeOH$  (2.2 mmol) were added to a 100 mL round-bottomed flask equipped with a magnetic stirring bar. The reaction mixture was stirred at room temperature for a half hour and the solvent was removed by rotary evaporation to produce the fluoride-releasing monomer (ternary zirconium–fluoride complex) 4. When antibacterial monomer 3 (2 mmol) was added to the above reaction mixture and under the same conditions, the product is the antibacterial fluoride-releasing (AF) monomer 5.

Analysis: HRMS (ESI) monomer 4 (negative ion mode, in MeOH) for  $C_{39}H_{47}F_2N_2O_{13}Zr$ ,  $[M - H]^-$ ,  $m/z$ : Found, 879.0; Calcd, 879.2099. Monomer 5 for  $C_{24}H_{40}NO_2^+$  (negative ion mode),  $m/z$ : Found, 879.0; (positive ion mode)  $[M3 - Br]^+$ ,  $m/z$ : 374.3051; Calcd, 374.3054.

Formulation of experimental composites: The compositions of experimental and control dental composites are shown in Table 1. Exp20, exp25, and exp30 contain 20, 25, and 39% of AF monomer 5 in the total monomer mixture. The monomers of control contain BisGMA/EBPADMA/HDDMA (40:40:20). The composites were formulated by blending the monomers 29%, silanized F-releasing glass filler 70%, and photoinitiators 1% in a Speed Mixer (Model CDAC 150FVZ, FlackTek, Inc.). The composite specimens were photopolymerized using an Optilux 501 dental curing light (Kerr Corp. Orange, CA) for 40 s on each surface.

Fluoride release and recharge: Fluoride release of both materials from cylindrical samples ( $4 \times 9$  mm (diameter  $\times$  height),  $n = 5$ ) in 3 mL of deionized water was analyzed daily for 30 days and then weekly for 65 days using an ion-selective electrode. Then the specimens were recharged by “60 Second Taste Gel” (containing 1.23 w/w% fluoride ion) for 1 min and rinsed with running deionized water for 30 s. Fluoride release after recharge was measured daily for 5 days and the recharge cycles were repeated three times.

Measurement of physical and mechanical properties: Cylindrical comprehensive strength specimens ( $4 \times 9$  mm (diameter  $\times$  height),  $n = 10$ ) and rectangular flexure strength specimens ( $25 \times 2 \times 2$  mm,  $n = 10$ ) were prepared, immersed in deionized water, stored at 37 °C for 24 h, and tested on the Instron 5566 testing machine with a crosshead speed of 1 mm/min. The water sorption and solubility test were conducted using disk specimens ( $15 \times 2$  mm (diameter  $\times$  height),  $n = 5$ ) according to ISO standard 4049.

Data analysis: The experimental data were analyzed using ANOVA and post hoc Tukey test with the significance level set at 95% ( $\alpha = 0.05$ ).

Cytotoxicity test: Human gingival fibroblasts were obtained from extracted molars from patients with healthy gingiva following informed consent, as prescribed in an approved IRB protocol. Gingival fibroblasts were maintained in MEM $\alpha$  containing 10% fetal calf serum (FCS) and 200 units/mL penicillin and 200  $\mu$ g/mL streptomycin. Cells were grown in 48-well plates for 24 h prior to exposure to the monomers. The growth media containing 0.1% DMSO were supplemented with  $10^{-4}$ ,  $10^{-5}$ ,  $10^{-6}$ , and  $10^{-7}$  M of synthesized monomers 2 and 4 were added to cells for 24 h. BisGMA served as a positive control for cytotoxicity. Cell survival was quantified using a Calcein AM labeling and BioTek Synergy 2 fluorescent multiwell plate reader.

## AUTHOR INFORMATION

### Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENTS

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